



Universitat de Lleida

# DEGREE CURRICULUM **LABORATORY METHODS IN PLANT BIOTECHNOLOGY**

Coordination: BASSIÉ , LUDOVIC RENÉ BERTIL

Academic year 2023-24

## Subject's general information

Subject name	LABORATORY METHODS IN PLANT BIOTECHNOLOGY			
Code	101629			
Semester	2nd Q(SEMESTER) CONTINUED EVALUATION			
Typology	Degree	Course	Character	Modality
	Bachelor's Degree in Biotechnology	4	OPTIONAL	Attendance-based
Course number of credits (ECTS)	6			
Type of activity, credits, and groups	Activity type	PRALAB	TEORIA	
	Number of credits	5	1	
	Number of groups	1	1	
Coordination	BASSIÉ , LUDOVIC RENÉ BERTIL			
Department	AGRICULTURAL AND FOREST SCIENCES AND ENGINEERING			
Teaching load distribution between lectures and independent student work	60 hores presencials 90 hores no presencials			
Important information on data processing	Consult <a href="#">this link</a> for more information.			
Language	Català 90% Anglès 10%			

Teaching staff	E-mail addresses	Credits taught by teacher	Office and hour of attention
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## Subject's extra information

The subject of **Lab Techniques in Plant Biotechnology (TLBP)** is an optional subject that is taught during the first semester of the fourth year of the Biotechnology Degree. The teaching load is 6 credits that are distributed between theoretical and practical classes with 1 and 5 credits respectively. The subject is structured in 4 theoretical sessions of 50 minutes, 3 sessions for the preparation of the mini-project (6 hours) and 50 hours of laboratory practices.

The TLBP subject is complementary to the subject of Plant Biotechnology (BTV). The tools used in plant biotechnology and the different transformation systems including the methods of integration of a transgene have already been explained in fully details in the subject of BTV.

To begin, we will see the methodology and strategy involved in the process of gene cloning and in subcloning stages. These techniques belong to the first crucial phase of plant genetic engineering.

The second group of techniques is devoted to methods for the analysis transgene expression, a fundamental post-transformation procedure to verify whether the gene of interest introduced into the host plant's genome is functional.

**Recommendation:** to have passed the subject Biotechnologia Vegetal (BTV)-101621

## Learning objectives

Objectives of knowledge:

The student that exceeds the subject must:

- have knowledge of elemental notions in the basic operation of a laboratory of biotechnology
- have complementary knowledge in genetic engineering strategies and adapted to the plant system
- have advanced knowledge in some relevant techniques of gene expression analysis

Objectives of capacity:

The student who exceeds the subject must be able:

- to know how to organize himself in the planning of the applied methodology
- to use efficient and reliable techniques for the analysis of gene expression
- to show a strong capacity of integration in a research team

## Competences

GC1 Being able to selectively search for and use sources of information necessary to achieve the training objectives.

GC3 Working in a team, with a multidisciplinary vision and with the ability to make a rational and efficient distribution of tasks among team members.

GC4 Knowing and adequately using the scientific and technical vocabulary of the different areas of Biotechnology.

GC5 Working in the laboratory applying criteria of quality and good practice.

GC6 Knowing and knowing how to use the specific software and databases in the different fields of biotechnology.

GC7 Using the scientific method to analyze data and design experimental strategies with biotechnological applications.

## Subject contents

**The theoretical classes** are based on the **practical and strategic aspect** in order to give the user the possibility to evolve quickly in learning the applied techniques and to provide him with advice and clear procedural explanations so that he can apply these methodologies of efficient way.

The program is structured in 4 parts:

-In the first part, theme-1 deals with the 'elementary notions' that must be applied and followed in the laboratory. The subject is addressed as a reminder way but with insistence on the essential and delicate points.

-Subsequently in topic 2, we explain the strategies involved at the different stages for isolating genes from both plant and bacterial origin. We see the stratagems and procedural tricks that can be used to initiate a gene cloning experiment and carry it out successfully. It is a key part for the preparation of the mini-project.

-In the third part (topic 3 and topic 4), In the third part we describe very briefly the methods of analysis to verify the expression of genes at transcriptional and translational level. Although they are classical methodologies, they are used systematically and are considered to provide very reliable results. They are techniques that must be applied to confirm results obtained by transcriptomic and proteomic approaches. We only focus on the critical points of the protocols, since these methods have been previously seen.

-To finish, we dedicate some sessions to the preparation of the mini-project (40% of the subject evaluation).

**Original ideas** are sought, within the framework of plant biotechnology, for **applications dedicated to agriculture or industry**. The aim is to describe the key points that are necessary for the generation of transgenic plants that express molecules of interest (enzymes and/or others) involved in the **engineering of a specific metabolic pathway** (endogenous or added).

## Topic 1. Generalities

Main safety measures in the laboratory: chemicals, ultraviolet light, general housekeeping- protective equipment, important safety messages, and emergency responses. Preparation and ownership of the common buffers and stock solutions. Sterilization by filtration and by autoclave. Maintenance of the laboratory notebook. The importance of following a protocol. Use of Equipment: General comments- Micropipettors- pH Meter- Autoclave operating procedures. Preparation of medium for E.coli bacterial culture. Work in sterile conditions. Working with E. coli: small scale cultures- Permanent storage.

## Topic 2. PCR Gene cloning and sub-cloning

### Gene cloning:

Choice of nucleic acid type for amplification: cDNA or genomic DNA. Primer design for the target sequence using databases. cDNA synthesis: the easiest way it is to perform the reaction in 2 steps. Amplification of the sequence of interest via PCR. PCR products: separation, extraction and purification. For TA-cloning: ligation reaction in a cloning vector. Competent cell transformation- Clone screening. Getting the sequence of interest via standard sequencing method (Sanger). How to design degenerate primers?

### Sub-cloning of insert(s) into an expression vector:

Adding via PCR appropriate restriction site at the extremities of the insert. Digestion of the vector and the insert with the respective enzymes. Fragment purification. Fragment quantification/ Ligation step/ Cell transformation.

### Gibson assembly- Gibson cloning

### Hairpin construct for producing a long dsRNA [iRNA strategy]

## Topic 3. RNA blot analysis

Reminder of working conditions with RNA. Methods for isolating total RNA: Trizol-like, silica-membrane based kits; LiCl precipitation. Checking RNA integrity. NaAc precipitation for concentrating RNA. Preparation of material for electrophoresis. Preparation of formaldehyde agarose gel. Preparation of RNA samples: denaturing buffer and thermal denaturation step. Electrophoretic separation and quality control. Wash of low salt strength followed by

transfer to a solid support. Fixing RNA by UV-crosslinking. DIG-labeled probe synthesis. Pre-hybridization and probe hybridization steps Sequential washes of variable stringency. DIG-labelled probe detection.

## **Topic 4. Western blotting**

Process of protein extraction. Bradford assay for protein quantification. Preparation of the solutions required for the gel and the transfer. PAGE-SDS electrophoresis: acrylamide gel preparation for denaturing condition (Laemmli method). Coomassie blue staining and destaining. Semi-dry transfer to PVDF membrane followed by blocking step. Types and properties of antibodies used for immunodetection. Incubation steps with primary and secondary antibodies. Detection of alkaline phosphatase activity (AP-conjugated secondary antibodies)/ detection of luminescence (HRP-conjugated antibody antibodies).

## Laboratory practices

Several technical procedures are organized.

The first one is devoted to protein analysis using the western blot method. Rice samples that overexpress the *gus* reporter gene controlled by the constitutive 35S CAMV promoter will be analyzed. All the stages of the methodology are carried out, from protein extraction from leaf tissue, through the preparation of the material to the final step by detecting the  $\beta$ -glucuronidase protein (GUS). In addition, as a complementary study, a Southern blot analysis is performed to identify the transgenic plant lines.

The second activity is dedicated to some key steps of gene cloning and sub-cloning procedures. High competent DH5 $\alpha$  E. Coli s cells are prepared according to the method of Hanahan. Efficiency is evaluated by using different quantities of a relevant plasmid and positive clones are screened by PCR after cell lysis.

The third activity is dedicated to RNA work. Student learns how to handle RNA samples by working in special conditions to preserve RNA integrity. Total RNA will be extracted from corn leaf tissue and the quality of the samples will be checked by electrophoresis.

It should be mentioned that the procedures of the different activities are done in a discontinuous manner; throughout the day steps of different protocols are performed in parallel, therefore optimizing time use. Time management is an important skill that the student must develop.

### **Day1: Protein isolation**

- Harvesting rice leaf samples, weighting and preservation in liquid nitrogen.
- Preparation of some buffers that are required for the two weeks of lab practices.
- Protein extraction procedure.
- Genomic DNA digestion from transgenic lines (*gus* transgene)

### **Day2: Western blot-I**

- Quantitative analysis of protein with the Bradford assay.
- Preparation of acrylamide gels (SDS-PAGE)
- Preparation of solid and liquid LB culture medium, with and without selection agent for E. coli DH5 $\alpha$  culture.

### **Day3: Western blot-II**

- Sample preparation for electrophoresis + gel running
- Assembly of the material (Mini-PROTEAN, Biorad)
- Blotting procedure + blocking step
- Plating-out DH5 $\alpha$  cells on non-selective LB medium.
- Preparation of transgenic seeds for germination

### **Day 4: Western blot-III + Southern blot**

- Washing steps

- Incubations with primary and secondary antibodies
- Colorimetric detection with alkaline phosphatase activity
- Interpretations of the results
- Mini culture of DH5 $\alpha$  cells in LB liquid media
- Southern blot: preparation of digested gDNA for electrophoresis and gel running.

## **Day 5: PAGE-SDS+ Southern blotting**

- Preparation of PAGE-SDS
- Preparation of TfbI and TfbII solutions (for competent cells preparation)
- Southern blot: ethidium bromide gel staining; denaturation; neutralization and transfer set up

## **Day 6: Isolation of RNA-I /Preparation of competent cells**

- Harvesting samples from rice leaf tissue; weighting and liquid nitrogen storage
- RNA extraction using LiCl precipitation
- Preparation of competent DH5 $\alpha$  cells (Hanahan protocol).

## **Day 7: Isolation of RNA-II / Transformation of competent cells**

- Continuation of the RNA isolation.
- Transformation of the competent DH5 $\alpha$  cells with two different plasmid conditions.
- Southern blot: pre-hybridization and probe hybridization

## **Day 8: Southern blot detection / PAGE SDS**

- Colony subcultures on selection medium (ampicillin)
- PAGE-SDS running (same conditions than day-3)
- Coomassie blue staining and destaining
- Southern blot: washing steps and procedure for probe detection

## **Day 9: Colony PCR screening + Qualitative RNA analysis**

- Cell lysis in TE buffer
- PCR: calculations and preparation of the mastermix for the detection of transformed bacterial clones with the plasmid of interest.
- Electrophoresis gel to check RNA integrity



**Day 10: PCR result**

- Preparation of agarose gels + electrophoresis to check gel to check PCR products
- Revision of all the results obtained during the different sessions.
- Questions and doubts

**Methodology**

Tipus d'activitat	Descripció	Activitat presencial alumne		Activitat no presencial alumne		Avaluació	Temps total
		Objectius	Hores	Treball alumne	Hores	Hores	Hores
<b>Lliçó magistral</b>	Classe magistral (Aula. Grup gran)	Explicació dels principals conceptes	6	Estudi: Conèixer, comprendre i sintetitzar coneixements			
<b>Problemes i casos</b>	Classe participativa (Aula. Grup gran )	Resolució de problemes i casos		Aprendre a resoldre problemes i casos			
<b>Seminari</b>	Classe participativa (Grup mitjà)	Realització d'activitats de discussió o aplicació		Resoldre problemes i casos. Discutir			
<b>Laboratori</b>	Pràctica de Laboratori (Grup mitjà)	Execució de la pràctica: comprendre fenòmens, mesurar...	50	Estudiar i realitzar Examen			
<b>Aula d'informàtica</b>	Pràctica d'aula d'informàtica (Grup mitjà )	Execució de la pràctica: comprendre fenòmens, mesurar...		Estudiar i Realitzar memòria			
<b>Pràctiques de camp</b>	Pràctica de camp (Grup mitjà )	Execució de la pràctica: comprendre fenòmens, mesurar...		Estudiar i Realitzar memòria			
<b>Visites</b>	Visita a explotacions o indústries	Realització de la visita		Estudiar i Realitzar memòria			
<b>Activitats dirigides</b>	Treball de l'alumne (individual o grup)	Orientar a l'alumne en el treball (en horari de tutories)	4	Realitzar un treball bibliogràfic, pràctic, etc.			
<b>Altres</b>							
<b>Totals</b>			60				

**Development plan**

See 'Contents' section

## Evaluation

### **-Laboratory report [60%]:**

The assessment of the report is based on the following points:

- General organization and coherence
- Details of the calculations and volumes used
- Explanation of issues
- Explanation of delicate steps
- Description and interpretation of the results obtained
- Participation, motivation and interaction among students

### **-Mini project of metabolic pathways engineering in plants [40%]:**

Project: find out an original idea, in the framework of plant biotechnology, for an application dedicated to agriculture or industry.

The objective is to describe the key points that are necessary for the generation of transgenic plants that express proteins of interest (enzymes and / or others) involved in the engineering of a specific metabolic pathway (endogenous or added).

Describe the following points with examples and technical information supported by the associated references.

- 1- Objective: Explain the purpose of your idea: selected plant species; study of the metabolic pathway (key enzymes)
2. Molecular construction/s: sequence analysis of the necessary gene(s) of interest; cloning strategy of the genes/s of interest; sub-cloning in appropriate vector(s) of expression.
3. The transformation method.
4. The key points for the regeneration of the transformed plants.
5. Expression analysis (RNA /protein): In order to characterize the transgenic plants, describe the most relevant method for transgene and/or endogenous gene expression.
- 6-Bibliography

## Bibliography

-Molecular Cloning- A Laboratory Manual. Vol 1,2,3. (Third Edition). 2001. J. Sambrook, DW. Russell. Cold Spring Harbor Laboratory Press, New York.

-Molecular Plant Biology Volume1, A practical approach. 2002. Philip M. Gilmartin and Chris Bowler. Oxford University Press.

[-www.protocol-online.org](http://www.protocol-online.org) (protocols i fòrums)